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Synthesis of 4,17-Diazasteroid Inhibitors of Human 5α-Reductase

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Abstract—The synthesis of the 17-aza isomer of finasteride is described. With the side chain amide group of the compound existing in the Z configuration the structure is similar to one of the two favored conformations of finasteride. A series of 4.17-diazasteroids was assayed against the isoenzymes of human 5α -reductase. Copyright © 1996 Elsevier Science Ltd

Introduction

Steroid 5α -reductase (EC 1.3.99.5) is the enzyme responsible for the NADPH-dependent conversion of testosterone to the more potent androgen dihydrotestosterone (DHT). There exist two isoenzymes¹⁻³ of 5α -reductase, type 1 and type 2, that are differentially expressed in various tissues such as skin, liver, prostate, seminal vesicle, and epididymis.⁴ The permissive role of DHT^{1.5} in benign prostatic hyperplasia, the most common neoplastic disease of aging men, is well established and DHT is also important in the viability of androgen-responsive cancer cells. These physiological roles for DHT have contributed to the interest in finding potent 5α -reductase inhibitors.

A number of 4-azasteroids, including finasteride 1, are potent type 2 inhibitors^{6,7} but 4,7β-dimethyl-4-azacholestan-3-one (2) and related 4-azacholestanes are selective type 1 inhibitors.8 Epristeride 3, an unsaturated 3-carboxysteroid, is a potent type 2 5α-reductase inhibitor⁹ and is active in models of prostatic cancer.¹⁰ To the same group of type 2 selective inhibitors belong 3-pyridyl-N-oxide steroids (e.g., 4)11 and 4-substituted-3-oxo- Δ^4 -steroids (e.g., 5). In addition, the latter compounds reduce the DHT-stimulated proliferation of androgen-sensitive Shionogi cells. Recent reports have described dual inhibition of both 5α-reductases by a series of 6-azasteroids (e.g., 6 and 7). 13,14 N-Amyl substituted 17 β -formamide 8^{15} has appeared to be one of the most potent inhibitors of human type 1 5α-reductase known so far. Nonsteroidal inhibitors (e.g., 9) of type 1 5α-reductase have also been described. 16 Probably all compounds mentioned above inhibit the enzyme because of their structural resemblance to the A-ring enol that is a presumed intermediate in the reduction of testosterone. 6,9,17 A common structural feature of the type 2 5α-reductase inhibitors is a 17β-carboxamide side chain substituted by a compact lipophilic residue.

Results and Discussion

In this communication we report the synthesis of finasteride isomer 18a and its derivatives with the amide nitrogen atom on the opposite side (compared with finasteride 1) of the 20-carbonyl group (i.e., 4,17-diazasteroids). Some 17-azasteroids (such as 10) have already been described as 'inverted' 5α-reductase inhibitors.¹⁸ These are, however, N-methyl-D-homo lactams with an amide side chain at the modified ring A. Our synthesis of 4,17-diazasteroids required prior preparation of 17-aza lactam 11,19 which was then reduced with lithium aluminum hydride to the secondary amine 12 followed by N-acylation with 3,3-dimethylbutyryl chloride. Oppenauer oxidation of the 3β-hydroxy compound 13 afforded α,β-unsaturated ketone 14. Further transformations were performed in a way similar to that described for finasteride 1 and its derivatives. 6 Compound 14 was oxidized with KMnO₄/ NaIO₄ to seco-acid 15 which was then subjected to ring closure with ammonia at high temperature to give the unsaturated lactam 16. The reduction of the C(5)—C(6) double bond was achieved by the method recently communicated by our group (NaBH₄/p-TsOH).²⁰ The addition of catalytic amounts of p-TsOH activates the enamide moiety to the borohydride reduction. The stereoselectivity (>85% of the 5α-H epimer 17a established by integration of signals in the ¹H NMR spectrum of the crude product) of reduction was as good as in the commonly used hydrogenation²¹ over a platinum catalyst. To complete the synthesis of the finasteride isomer 18a 1-dehydrogenation was performed by silylation-mediated [bis(trimethylsilyl)trifluoroacetamide was used DDQ oxidation. 22,23 4-Methyl derivatives of 4,17-diaza lactams (17b) and 18b) were also obtained by CH₃I/NaH methylation.

The conformation of the amide side chain needs some comment. Molecular mechanics calculations (MM + force field)²⁴ suggest that the finasteride 17-aza isomer

Scheme 1.

18a preferentially exists in the Z configuration with the torsion angle C(13)—N—C(20)—O close to zero (Fig. 1). Significant contribution of this configuration was proved by ¹H NMR spectra of compounds 13-18 where 12 β -H is strongly deshielded (δ 3.0–3.1 ppm)²⁵ by the nearby amide carbonyl group. The alternative E configuration (Fig. 2) is less stable by 5.7 kcal/mol and is virtually without significance. The preference of the Z configuration in 18a was also confirmed by ¹H NOE difference spectroscopy. Upon irradiation of the side chain methylene protons, a 2.2% enhancement of the 16β-H signal at δ 3.53 and a 1.0% enhancement of the 16α -H signal at δ 3.42 were observed. There was also enhancement at δ 1.05 (t-butyl protons), but no detectable NOE effect was found for 12β-H. In the ¹H 2D NOESY spectrum of 18a two cross-peaks between the methylene group and both protons at C-(16) were present, whereas no spatial interaction of the side chain protons with 12β-H was found. This excludes the existence of E configuration with a short distance between 12β-H and one of the methylene group protons (about 2.05 Å).

The preference of the Z configuration of the amide moiety was found in finasteride 1 [torsion angle O-C(20)-N-C(t-butyl) amounts 8.3° and other unsymmetrically substituted side chain amides. There are two favored, almost equally populated conformations about the C(17)-C(20) bond in a series of 20-carboxylic amides (see Table 1). In the case of

unsubstituted and monosubstituted amides the amide plane is deviated from the ring D plane by 20–25° for one conformation and by 110–115° for another. The amide group of disubstituted amides may lie in the ring D plane or be perpendicular to it.

As shown in Figure 3, the preferred side chain conformation of the 17-aza isomer is similar to one of the favored finasteride conformations. However, there is more restriction in the side chain rotation in the 17-aza isomer than in the case of finasteride itself.

A series of 4,17-diazasteroids was assayed against both type 1 and type 2 5α-reductase. The methods employed for determining enzyme inhibition data (Table 2) were as previously described.¹³

The finasteride 17-aza isomer **18a** proved to be quite a potent inhibitor of type 2 5α -reductase, although it is less active than finasteride **1** and its congeners. It seems that the conformation found in **18a** is not optimal for the inhibitory activity. Compound **17b** was found to be an inhibitor of both type 1 and type 2 5α -reductases. The results of assays performed show that 4-methylation increases activity in the case of compound **17**, but lowers the inhibition of the type 2 enzyme by the Δ^1 derivative **18**. Compound **16** with a C(5)—C(6) double bond shows only moderate inhibition of human type 2 5α -reductase activity. These results are consistent with general rules established for

Scheme 2.

the 4-azasteroid derivatives.⁶ The 4-methyl azasteroids are more active than the unsubstituted analogues when the A ring is saturated, the reverse is true for the Δ^1 derivatives. Unsaturation at C5(6) resulted in reduced 5α -reductase inhibitory activity.

We are pursuing synthetic chemistry towards new azasteroid inhibitors of 5α -reductase.

Experimental

Melting points were determined on a Köffler apparatus of the Boetius type and were uncorrected. NMR spectra were taken with a Bruker AC 200F and a Varian UNITYplus-500 spectrometers using CDCl₃

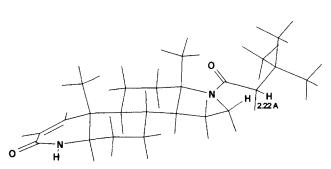


Figure 1. Configuration Z of the finasteride 17-aza isomer 18a with the shortest H—H contact shown.

solutions with TMS as the internal standard. IR spectra were recorded on a Specord 75 IR spectrophotometer as chloroform solutions. Mass spectra were obtained at 70 eV with an AMD-604 spectrometer. The reaction products were isolated by column chromatography performed on 70–230 or 230–400 mesh silica gel (Merck). Thin-layer chromatograms were developed on aluminum TLC sheets precoated with silica gel F_{254} and visualized with 50% sulfuric acid after heating. All solvents were dried and freshly distilled prior to use.

Type 1 and type 2 recombinant human 5α -reductase assays were carried out as previously described.¹³

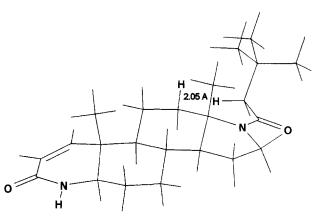


Figure 2. Configuration E of the finasteride 17-aza isomer 18a with the shortest H-H contact shown.

Table 1. Least energy conformations of 5α -reductase inhibitors

	Steric energy of conformation (kcal/mol); torsion angle C(13)—C(17)—C(20)—O		$\Delta E_{\text{(I-II)}}$ (kcal/mol)	Human prostatic 5α-reductase ⁶ IC ₅₀ /IC ₅₀ Fin.
CONR ₁ R ₂	O _C N _{R₂}	R ₂ N _C O		
$R_1 = R_2 = H$ $R_1 = C_2H_5$; $R_2 = H$ $R_1 = C(CH_3)_3$; $R_2 = H$ (Fin.) $R_1 = C(CH_3)_2CH_2C(CH_3)_3$; $R_2 = H$ $R_1 = R_2 = C_2H_5$ $R_1 = R_2 = CH(CH_3)_2$ Finasteride 17-aza isomer	27.9; -19.9° 29.1; -24.1° 32.9; -25.9° 42.9; -23.1° 37.5; -1.7° 40.5; 0.6° 35.7; -0.9° °	27.5; 115.8° 28.3; 112.4° 31.6; 115.4° 41.9; 111.5° 37.3; 93.5° 40.6; 90.7° 41.4; 177° a	0.4 0.8 1.3 1.0 0.2 -0.1 -5.7	151 27 1 2.2 11 2.9 16 ^b

^{*}Torsion angle C(13)—N—C(20)—O.

17-(3,3-Dimethylbutyryl)-17-azaandrost-5-en-3β-ol (13). To a suspension of LiAlH₄ (550 mg, 14.5 mmol) in anhydrous dioxane (100 mL), lactam 11 (1.03 g; 3.1 mmol) was added and the reaction mixture was refluxed for 2 days. The reaction was carefully quenched with stoichiometric amount of water (1.05 mL, 58.3 mmol) and to the resulting solution of amine 12, 3,3-dimethylbutyryl chloride (1.5 mL, 10.7 mmol) was added dropwise. The reaction mixture was stirred

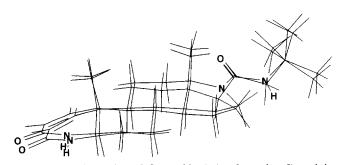


Figure 3. RMS overlay of finasteride 1 (conformation I) and its 17-aza isomer 18a.

overnight at room temperature, all inorganic material was filtered off, and the solvent was removed in vacuo. The crude product was purified by silica gel column chromatography. Amide 13 (840 mg; 72%) was eluted with benzene:ethyl acetate (3:1) preceded by small amounts of its O-3,3-dimethylbutyryl derivative (18 mg; 1%). Amide 13; mp 214-217°C (hexane:methylene chloride); IR (CHCl₃ cm⁻¹): 3610, 3400, 1609, 1403, 1045; ¹H NMR: δ 5.35 (m, 1H, 6-H), 3.33–3.66 (m, 3H, 3α -H and 16-H), 3.06 (m, 1H, 12 β -H), 2.06 and 2.10 (AB system, J = 13.9 Hz, 2H, C_{H_2} -t-Bu), 1.15 (s, 3H, 18-H), 1.05 (s, 9H, t-Bu), 1.02 (s, 3H, 19-H); ¹³C NMR: δ 171.0 (C), 141.4 (C), 120.7 (CH), 71.6 (CH), 62.9 (C), 54.8 (CH), 50.2 (CH), 47.8 (CH₂), 47.2 (CH₂), 42.2 (CH₂), 37.4 (CH₂), 37.2 (CH₂), 36.6 (C), 31.6 (C), 31.44 (CH), 31.35 (C), 30.0 ($3 \times \text{CH}_3$), 29.7 (CH₂), 24.6(CH₂), 21.9 (CH₂), 19.3 (CH₂), 15.6 (CH₂); MS: m/z (rel. int.) 373 (11), 358 (16), 317 (17), 302 (15), 284 (13), 274 (7), 260 (100).

17-(3,3-Dimethylbutyryl)-17-azaandrost-4-en-3-one (14). From a solution of amide 13 (820 mg, 2.2 mmol) in toluene (30 mL) and cyclohexanone (5 mL), ca. 10 mL

Table 2. Inhibition of recombinant type 1 and type 2 human 5α -reductase by 4,17-diazasteroids (IC_{s0}s with the standard deviations from the fitted lines in parentheses)

Compound no.	Type 1 5α-reductase IC ₅₀ (nM)	Type 2 5α-reductase IC ₅₀ (nM)	
16	~7000	52.0 (±7.8)	
17a	$2200 \ (\pm 140)$	$40.1~(\pm 2.8)$	
17b	$28.0\ (\pm 2.1)$	$3.6 (\pm 0.3)$	
18a	$765 (\pm 70)$	$10.3~(\pm 1.1)$	
18b	$477\ (\pm 29)$	$174(\pm 42)$	
4-MA ^a	$6.4~(\pm 0.2)$	$0.4 (\pm 0.04)$	

 $[^]aN_iN_i$ -Diethyl-3-oxo-4-methyl-4-aza-5 α -androstane-17 β -carboxamide (4-MA) was used as a standard reference.

^bCalculated on the basis of type 2 recombinant human 5α-reductase assays.

of the solvent was distilled off and then aluminum tri-iso-propoxide (400 mg, 1.96 mmol) in anhydrous toluene (5 mL) was added dropwise. The reaction mixture was gently boiled during 1 h, while another 10 mL of the solvent was distilled off. After cooling, 20 mL of saturated solution of sodium potassium tartrate was added, and the organic solvents were removed by steam distillation. The reaction mixture was diluted with water, extracted with chloroform, and the dried (anhydrous MgSO₄) extract was evaporated under reduced pressure. The crude product was purified by silica gel column chromatography (elution with 25% ethyl acetate:benzene). Yield of 14: 675 mg (83%); mp 152–155 °C (hexane:methylene chloride); IR (CHCl₃, cm⁻¹): 1700, 1610, 1402, 1196; ¹H NMR: δ 5.74 (m, 1H, 4-H), 3.35-3.63 (m, 2H, 16-H), 3.04 (m, 1H, 12β -H), 2.06 and 2.10 (AB system, J = 13.9 Hz, 2H, C_{H_2} -t-Bu), 1.19 (s, 3H, 19-H), 1.17 (s, 3H, 18-H), 1.05 (s, 9H, t-Bu); 13 C NMR: δ 199.3 (C), 170.3 (2×C), 124.0 (CH), 62.7 (C), 54.0 (CH), 53.6 (CH), 47.7 (CH₂), 47.1 (CH₂), 38.5 (C), 37.2 (CH₂), 35.6 (CH₂), 34.8 (CH), 33.8 (CH₂), 32.5 (CH₂), 31.44 (CH₂), 31.38 (C), 29.9 $(3 \times CH_3)$, 24.4 (CH₂), 21.8 (CH₂), 17.1 (CH₃), 15.6 (CH_3) ; MS: m/z (rel. int.) 371 (16), 356 (15), 315 (33), 301 (18), 272 (4), 258 (100); HRMS: m/z calcd for C₂₄H₃₇O₂N [M]⁺, 371.2824. Found: 371.2822.

17-(3,3-Dimethylbutyryl)-4-nor-3,5-seco-5-oxo-17-azaandrostan-3-oic acid (15). To a stirred solution of compound 14 (660 mg, 1.78 mmol) in tert-butanol (10 mL) was added 1.5 mL of hot aqueous solution of sodium carbonate (0.3 g). The reaction mixture was heated at reflux,²⁶ while a warm aqueous solution (1.5 mL) of NaIO₄ (3.2 g, 14.96 mmol) and KMnO₄ (40 mg, 0.25 mmol) was added portionwise. After addition of the last portion of the oxidizing agents, the reaction was refluxed for another 1 h. All inorganic material was filtered off, and washed with water and chloroform. The filtrate was acidified with diluted hydrochloric acid and extracted with chloroform. The solvent was removed from the dried (anhydrous MgSO₄) extract in vacuo and the residue was purified by silica gel column chromatography. seco-Acid 15 (497 mg, 71%) was eluted with benzene:ethyl acetate (1:9); mp 58-60° C (hexane:benzene); IR (CHCl₃, cm⁻¹): 1692, 1614, 1403, 1201; ¹H NMR: δ 7.55 (bs, 1H, COOH), 3.35–3.65 (m, 2H, 16-H), 3.04 (m, 1H, 12β-H), 2.54 (m, 1H, 2-H), 2.15 and 2.19 (AB system, J = 13.9 Hz, 2H, CH₂-t-Bu), 1.20 (s, 3H, 19-H), 1.12 (s, 3H, 18-H), 1.06 (s, 9H, t-Bu); ¹³C NMR: δ 200.3 (C), 178.1 (C), 171.8 (C), 63.5 (C), 54.0 (CH), 50.2 (C), 47.9 (CH₂), 47.7 (CH), 46.8 (CH₂), 37.6 (CH₂), 36.9 (CH₂), 34.1 (CH), 31.8 (C), 30.6 (CH₂), 30.0 ($3 \times \text{CH}_3$), 29.1 (CH₂), 28.9 (CH₂), 24.3 (CH₂), 22.2 (CH₂), 20.1 (CH₃), 15.8 (CH₃); MS: m/z (rel. int.) 391 (12), 376 (15), 335 (23), 320 (4), 303 (9), 278 (100); HRMS: m/z calcd for $C_{23}H_{37}O_4N$ [M]+, 391.2723. Found: 391.2723.

17-(3,3-Dimethylbutyryl)-4,17-diazaandrost-5-en-3-one (16). To a stirred suspension of seco-acid 15 (440 mg, 1.13 mmol) in ethylene glycol (6 mL) liquid ammonia (2 mL) was added dropwise. The homogeneous

solution was then gradually heated (3 °C/min) to 180 °C and held at 180 °C for 15 min. After cooling, the mixture was acidified with diluted hydrochloric acid and extracted with chloroform. The extract was washed three times with water, dried (anhydrous MgSO₄), and evaporated in vacuo. The crude product 16 was purified by silica gel column chromatography (elution with 3% methanol:chloroform). Yield 326 mg (78%); mp 222-225 °C (benzene); IR (CHCl₃ cm⁻¹): 3408, 1662, 1631, 1416, 1210; ¹H NMR: δ 8.58 (s, 1H, NH), 4.93 (m, 1H, 6-H), 3.32-3.60 (m, 2H, 16-H), 3.06 (m, 1H, 12β-H), 2.47 (m, 2H, 2-H), 2.06 and 2.10 (AB system, J = 13.8 Hz, 2H, CH_2 -t-Bu), 1.17 (s, 3H, 19-H), 1.10 (s, 3H, 18-H), 1.05 (s, 9H, t-Bu); 13 C NMR: δ 171.1 (C), 170.0 (C), 140.2 (C), 102.8 (CH), 68.4 (C), 62.8 (C), 54.4 (CH), 47.9 (CH), 47.7 (CH₂), 47.1 (CH₂), 37.0 (CH₂), 34.1 (C), 31.4 (CH₂), 31.0 (CH), 29.9 $(3 \times CH_1)$, 29.1 (CH₂), 28.2 (CH₂), 24.4 (CH₂), 21.6 (CH_2) , 18.5 (CH_3) , 15.6 (CH_3) ; MS: m/z (rel. int.) 372 (18), 357 (10), 316 (18), 301 (42), 273 (6), 259 (100); HRMS: m/z calcd for $C_{23}H_{36}O_2N_2$ [M]⁺, 372.2777. Found: 372.2777.

17-(3,3-Dimethylbutyryl)-4,17-diaza-5α-androstan-3-one (17a). A stirred solution of diazasteroid 16 (180 mg, 0.48 mmol) in THF (20 mL) was treated with concd sulfuric acid (0.08 mL, 1.52 mmol) and NaBH₄ (600 mg, 15.8 mmol). Borohydride was added portionwise during 5 h and stirring was continued for another 1 h. The reaction was quenched with water and extracted with chloroform. The solvent was evaporated from the dried (anhydrous MgSO₄) extract and the residue was chromatographed on a silica gel column. The pure product 17a (96 mg; 53%) was eluted with 3% methanol:chloroform preceded by fraction (23 containing 17a and its 5β-epimer; mp 285-288 °C (benzene); IR (CHCl₃, cm⁻¹): 3406, 1640, 1621, 1396; ¹H NMR: δ 6.00 (bs, 1H, NH), 3.32–3.60 (m, 2H, 16-H), 3.06 (m, 2H, 5α -H and 12β -H), 2.40 (m, 2H, 2-H), 2.06 and 2.10 (AB system, J = 13.8 Hz, 2H, CH₂-t-Bu), 1.13 (s, 3H, 18-H), 1.05 (s, 9H, t-Bu), 0.90 (s, 3H, 19-H); ¹³C NMR: δ 172.4 (C), 170.9 (C), 63.1 (C), 60.6 (CH), 54.0 (CH), 51.3 (CH), 47.7 (CH₂), 47.1 (CH_2) , 37.3 (CH_2) , 35.6 (C), 34.3 (CH), 33.2 (CH_2) , 31.4 (C), 29.9 ($3 \times \text{CH}_3$), 29.1 (CH₂), 28.5 (CH₂), 27.0 (CH₂), 24.3 (CH₂), 21.9 (CH₂), 15.7 (CH₃), 11.2 (CH₃); MS: m/z (rel. int.) 374 (10), 359 (11), 318 (28), 303 (8), 275 (4), 261 (100); HRMS: m/z calcd for $C_{23}H_{38}O_2N_2$ [M]⁺, 374.2933. Found: 374.2931.

17-(3,3-Dimethylbutyryl)-4,17-diaza-5 α -androst-1-en-3-one (18a). A stirred solution of compound 17a (87 mg, 0.23 mmol) in anhydrous dioxane (1 mL) was treated with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (60 mg; 0.26 mmol) and bis(trimethylsilyl)trifluoroace-tamide (0.3 mL, 1.11 mmol) under N₂. The reaction mixture was stirred at room temperature for 4 h and then at 110 °C for 18 h. The resulting dark solution was diluted with methylene chloride, washed with 2% aqueous sodium bisulfite solution, 2 N hydrochloric acid, water, dried (anhydrous MgSO₄) and evaporated.

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The crude product 18a was purified by silica gel column chromatography (elution with 3% methanol: chloroform). Yield 66 mg (76%), mp 245-248°C (benzene); IR (CHCl₃, cm⁻¹): 3432, 1671, 1624, 1412; ¹H NMR: δ 6.79 (d, J = 10.0 Hz, 1H, 1-H), 5.90 (broad signal, 1H, NH), 5.81 (dd, J = 10.0 Hz, 1.9 Hz, 1H, 2-H), 3.53 (t, J = 9.5 Hz, 1H, 16 β -H), 3.42 (ddd, J = 9.7Hz, 9.5 Hz, 7.5 Hz, 1H, 16α -H), 3.35 (dd, J = 11.5, 4.6 Hz, 1H, 5α -H), 3.09 (m, 1H, 12β -H), 2.06 and 2.11 (AB system, J = 13.7 Hz, 2H, CH_2 -t-Bu), 1.14 (s, 3H, 18-H), 1.05 (s, 9H, t-Bu), 0.98 (s, 3H, 19–H); 13 C NMR: δ 171.1 (C), 166.6 (C), 150.7 (CH), 123.1 (CH), 63.1 (C), 59.6 (CH), 54.0 (CH), 47.7 (CH₂ and CH), 47.2 (CH₂), 39.4 (C), 37.4 (CH₂), 34.6 (CH), 31.4 (C), 30.0 $(3 \times CH_3)$, 29.1 (CH₂), 25.8 (CH₂), 24.4 (CH₂), 22.1 (CH_2) , 15.8 (CH_3) , 11.9 (CH_3) ; MS: m/z (rel. int.) 372 (10), 357 (12), 316 (31), 301 (24), 273 (4), 259 (100); HRMS: m/z calcd for $C_{23}H_{36}O_2N_2$ [M]⁺, 372.2777. Found: 372.2777; Anal. calcd for $C_{23}H_{36}O_2N_2$: C, 74.15; H, 9.74; N, 7.52. Found: C, 74.04; H, 9.79; N, 7.56.

17-(3,3-Dimethylbutyryl)-4-methyl-4,17-diaza-5α-androstan-3-one (17b) and its 1-dehydro derivative 18b. solution of diazasteroid (17a or 18a; 37 mg, 0.1 mmol) in anhydrous DMF (3 mL) was treated with NaH (60% suspension in oil; 10 mg, 0.25 mmol). The reaction mixture was stirred at room temperature for 15 min, then methyl iodide (0.2 mL, 3.21 mmol) was added and stirring was continued for 1 h at 40 °C. The mixture was poured into water, acidified and extracted with chloroform. The crude products (17b or 18b) were purified by silica gel column chromatography (elution with 3-4% methanol:chloroform). Yield of 17b 35 mg (92%), mp 199-203 °C (hexane:methylene chloride); ÎR (CHCl₃, cm⁻¹): 1623, 1431; ¹H NMR: 3.32–3.60 (m, 2H, 16-H), 3.05 (m, 2H, 5α -H and 12β -H), 2.93 (m, 3H, N-CH₃), 2.45 (m, 2H, 2-H), 2.06 and 2.10 (AB system, J = 13.85 Hz, 2H, CH_2 -t-Bu), 1.13 (s, 3H, 18-H), 1.05 (s, 9H, t-Bu), 0.89 (s, 3H, 19-H); 13 C NMR: δ 171.0 (C), 170.7 (C), 65.6 (CH), 63.1 (C), 54.0 (CH), 51.9 (CH), 47.8 (CH₂), 47.1 (CH₂), 37.4 (CH₂), 36.4 (C), 33.6 (CH), 32.8 (CH₂), 31.4 (C), 30.0 ($3 \times$ CH₃), 29.8 (CH₂), 29.1 (CH₃), 29.0 (CH₂), 25.2 (CH₂), 24.3 (CH_2) , 21.9 (CH_2) , 15.7 (CH_3) , 12.2 (CH_3) ; MS: m/z(rel. int.) 388 (12), 373 (13), 332 (28), 317 (10), 289 (4), 275 (100). HRMS: m/z calcd for $C_{24}H_{40}O_2N_2$ [M]⁺, 388.3090. Found: 388.3091. Yield of **18b** 34 mg (89%); mp 169-172 °C (hexane:methylene chloride); IR (CHCl₃, cm⁻¹): 1658, 1620, 1417, 1203, 1102; ¹H NMR: δ 6.66 (d, J = 9.8 Hz, 1H, 1-H), 5.83 (d, J = 9.8 Hz, 1H, 2-H), 3.30-3.60 (m, 3H, 5α -H and 16-H), 3.05 (m, 1H, 12β-H), 2.93 (s, 3H, N-CH₃), 2.06 and 2.10 (AB system, J = 13.8 Hz, 2H, CH_2 -t-Bu), 1.11 (s, 3H, 18-H), 1.01 (s, 9H, t-Bu), 0.89 (s, 3H, 19-H); 13 C NMR: δ 171.0 (C), 165.5 (C), 140.4 (CH), 123.2 (CH), 63.6 (CH), 62.9 (C), 53.8 (CH), 47.8 (CH), 47.6 (CH₂), 47.0 (CH₂), 39.5 (C), 37.3 (CH₂), 33.9 (CH), 31.4 (C), 29.9 $(3 \times CH_3)$, 29.5 (CH₂), 27.6 (CH₃), 24.3 (CH₂), 24.2 (CH₂), 22.0 (CH₂), 15.7 (CH₃), 12.0 (CH₃); MS: m/z (rel. int.) 386 (14), 371 (12), 330 (29), 315 (29), 287 (5), 273 (100); HRMS: m/z calcd for $C_{24}H_{38}O_2N_2$ [M]⁺, 386.2933. Found: 386.2933.

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- 25. The signal was unequivocally assigned to the 12β -proton by considering its coupling pattern and connectivities established by a 1H 2D COSY spectrum, as well as the through-space proximity data obtained from a 1H 2D NOESY spectrum.
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